Review

# **Prevention of Vitamin D Deficiency and Osteoporosis**

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# Abstract

It is well-known that vitamin D status impacts on the mineralization of the skeleton, bone turnover rate, and the occurrence of fractures. Numerous studies have shown a direct relationship between a low serum level of vitamin D and a low bone mineral density (BMD) and an increased risk of non-vertebral and hip fractures. Importantly, vitamin D status is essential for the generation of maximal bone mass and along with other risk factors can largely contribute the development of osteoporosis and other bone metabolic diseases, such as osteomalacia and rickets in children. Among bone metabolic disorders, osteoporosis is the most common chronic skeletal disease with high morbidity and mortality rates and its healthcare costs impose a significant economic burden. The effective prevention of vitamin D deficiency and osteoporosis can be achieved with the timely identification of individuals with an increased risk for vitamin D deficiency and subsequent osteoporosis was presented on the basis of evolutionary medicine principles. Moreover, we have highlighted the possibility of using selected plants as natural sources of vitamin D for reducing the incidence of vitamin D deficiency, and efficiency. Several other plant-derived compounds, such as phytoestrogens, flavonoids, and

polyphenols, which exhibited protective effects on bone metabolism, were also explored for their potential use for nutritional prevention of osteoporosis.

**KeyWords**: vitamin D, vitamin D deficiency, osteoporosis, bone metabolism, cholecalciferol, ergocalciferol, VDR.

### 1. Introduction

According to the current knowledge, vitamin D status impacts on the mineralization of the skeleton, bone turnover rate, and the occurrence of fractures. Based on serum 25hydroxyvitamin D (250HD) levels, the most abundant metabolite of vitamin D in the circulation, the global consensus recommends the following classification of vitamin D status: sufficiency, >50 nmol/l; insufficiency, 30-50 nmol/l; deficiency, <30 nmol/l [1]. A study showed that patients with serum 25-(OH)D levels lower than 50 nmol/l had accelerated bone turnover, bone loss, and feasible mineralization defects [2]. Other studies have also shown that low 25(OH)D serum level is directly related to a low bone mineral density (BMD) and an increased risk of non-vertebral and hip fractures [3] [4] [5]. The optimal serum concentration of vitamin D is important for the generation of maximal bone mass [6], which is decreasing with age in both men and women leading to a bone loss. In old age, osteoporosis usually occurs, as a result of bone loss, unless timely treatment has been applied. In particular, osteoporosis is developed in elderly or postmenopausal women since they experience a prevalently pronounced increase in bone loss [2] [7]. However, osteoporosis can also occur in men with osteoporotic fractures affecting one in eight men at the age of over 50 [8]. The reduced bone mass and bone architecture disruption eventually leading to fragility fractures are major features of osteoporosis. Osteoporosis-associated loss of bone is a gradual process that usually occurs over the years and is usually silent until the bones are so frail that a fracture occurs [9]. Among bone metabolic disorders, osteoporosis is the most common chronic skeletal disease with high morbidity and mortality rates [8] [10] and its healthcare costs impose a significant economic burden [11]. Several epidemiological studies investigated the relationship between vitamin D deficiency and the incidence of osteoporotic fractures [12] [13]. Of note, in 2000, nine million osteoporotic fractures were registered globally [14] and the projection remains consistent [15]. In particular, the Longitudinal Aging Study Amsterdam presented data indicating that people with serum concentration of 25(OH)D lower than 30 nmol/l have a higher incidence of osteoporotic fractures [12]. Furthermore, many interventional studies have revealed that vitamin D supplements with or without calcium can reduce the fracture incidence in older individuals [16] [17]. Besides, fractures may occur in patients with osteomalacia and in children suffering from rickets. These conditions occur only when vitamin D deficiency is severe (i.e., serum level of 25-hydroxyvitamin D is below 15 nmol/l) [18]. Therefore, vitamin D deficiency is an important research target aimed at improving its prevention, and consequently, the cost-effectiveness of care of patients with osteoporosis. In this review, the rationale for identifying those at risk for vitamin D deficiency and subsequent osteoporosis was presented on the basis of evolutionary medicine principles. Osteomalacia and rickets will not be discussed here in detail. Moreover, we highlighted the possibility of using the selected plants as natural sources of vitamin D for reducing the incidence of vitamin D deficiency. Additionally, several other plant-derived compounds, such as phytoestrogens, flavonoids, and

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polyphenols, which exhibited protective effects on bone metabolism, were also explored for their potential use for nutritional prevention of osteoporosis.

# 2. Vitamin D Biology and Pathophysiologic Pathways of Bone Loss and Mineralization Defects in Vitamin D Deficiency

It is well known that sufficient quantities of minerals, such as calcium and phosphorus promote healthy bone tissue development in both animals and humans. Adequate levels of these minerals are ensured by both sufficient dietary intake and metabolism, one of the most important regulators of which is vitamin D [19]. There are two forms of vitamin D, such as vitamin D<sub>2</sub> or ergocalciferol and vitamin D<sub>3</sub> or cholecalciferol, which differ in their chemical side chains. Vitamin D<sub>3</sub> is endogenously synthesized from its precursor 7-dehydrocholesterol in skin cells upon ultraviolet (UV) light exposure; whereas,  $D_2$  is produced in yeast and plants, therefore, can be ingested with some nutrients or supplements [19]. Vitamin D<sub>3</sub> is also present in some nutrients, such as sunflower oil, fish, and rainbow trout [20]. Vitamin D metabolism that maintains serum calcium and phosphorus levels in physiologically acceptable ranges and provides skeleton mineralization was described in detail by Holick [21]. Vitamin D<sub>3</sub> is converted in the liver to 25-hydroxycholecalciferol D and further hydroxylated by the kidneys forming 1,25-dihydroxycholecalciferol D [25(OH)<sub>2</sub>D], the biologically active compound of vitamin D. Vitamin D<sub>2</sub> is metabolized to 25-hydroxyergocalciferol. Vitamin D metabolites, 25hydroxycholecalciferol and 25-hydroxyergocalciferol, also called 25-hydroxyvitamin D or 25(OH)D, are measured in serum to determine a patient's vitamin D status. Activated vitamin D contributes to the maintenance of serum calcium levels by increasing calcium intestinal absorption and by stimulating osteoclastic bone resorption [9]. Besides, activated vitamin D promotes intestinal absorption of phosphate. Thus, bone tissue homeorhesis can be maintained by the balance of vitamin  $D_3$  endogenous production and dietary consumption of vitamin  $D_2$ .

In the case of vitamin D deficiency, the level of 1,25(OH)2D may decline and a smaller amount of calcium will be available for bone mineralization. The parathyroid hormone (PTH) concentration will increase (secondary hyperparathyroidism), accelerating the hydroxylation of 25(OH)D to 1,25(OH)2D in the kidney. The elevated serum PTH promotes bone turnover and increased bone resorption leading to bone loss [22] and these processes contribute to the pathogenesis of osteoporosis. In this way, PTH stimulates the activity of osteoblasts, which, in turn, accelerate the transition of pre-osteoclasts into mature osteoclasts. Osteoclasts dissolve the mineralized collagen matrix of bone, triggering osteopenia and osteoporosis and increasing the risk of fracture. The increased serum PTH levels are associated with common in postmenopausal women vitamin D deficiency [23]. Noteworthy, apart from vitamin D deficiency, the pathophysiology osteoporosis involves the interaction between many factors, including low peak BMD and sex hormone deficiency among others [24]. In addition, an epidemiological study of 237 women with post-menopausal osteoporosis also showed that a low level of vitamin D is associated with some determinants of osteoporosis and falls [25].

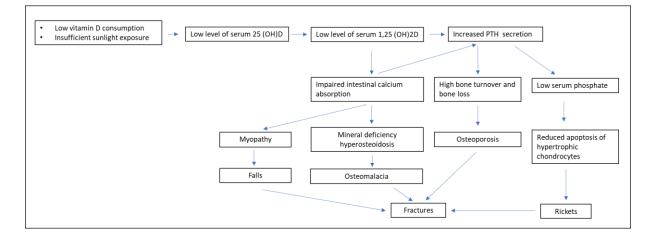
In contrast to the frequently-presented association between vitamin D deficiency and osteoporosis, an early study demonstrated that there was no relationship between serum concentration of 25-hydroxyvitamin D and BMD for the spine, hip, and total body skeleton [26]. This study included 262 healthy urban Chinese women aged between 40 and 72 years, which were randomly selected in Taipei city. The authors concluded that since reduced bioavailability of vitamin D did not determine BMD, hence, it is insignificant in osteoporosis

development, in the free-living urban Chinese population. Of note, BMD is the major predictor of osteoporosis and fracture risk, particularly in postmenopausal women [27].

When vitamin D deficiency is persistent and severe, osteomalacia can be developed. Osteomalacia is associated with a decrease in bone volume and excessive osteoid accumulation resulting in bone mineralization defects in adults and adolescents [28] and always goes alongside rickets in children. Rickets is a disease of defective chondrocyte differentiation, defective mineralization of the growth plate, and defective osteoid mineralization, which is caused by vitamin D deficiency and/or low calcium intake in children [1]. As a consequence of secondary hyperparathyroidism, hypophosphataemia resulting in decreased apoptosis of hypertrophic chondrocytes in the growth plate and diminished mineralization of primary spongiosa in the metaphysis (new bone) is the underlying pathomechanism of rickets of all forms [29].

The effect of vitamin D deficiency on bone leading to the development of osteoporosis, osteomalacia, rickets, and fractures was presented in **Figure 1**.

**Figure 1.** Schematic representation of the effect of vitamin D deficiency on bone leading to the development of osteoporosis, osteomalacia, rickets, muscle weakness, and fractures.



### 3. Identifying Those at Risk for Vitamin D Deficiency and Osteoporosis

The problem of vitamin D deficiency can undoubtedly be solved, but approaches to optimize vitamin D status may require further improvements [30]. In this respect, using the principles of evolutionary biology [31] may help to identify those at risk for vitamin D deficiency. According to this paradigm, the evolutionarily developed genetic traits determining vitamin D status under certain environmental conditions should be taken into account in the development of risk assessment strategies. For example, a recent study explored the relationship between gene polymorphisms influencing bone tissue metabolism, latitude, and food habits [32]. Interestingly, their study indicated that the geographic localization or latitude of the population per se was not a prominent factor affecting vitamin D status. Apparently, evolutionary acquired vitamin D receptor (*VDR*) variants, such as *CC\*Fok1*, *CT\*Fok1*, and *GA\*Bsml*, were found to determine healthy bone mineral metabolism in the studied population, despite living in areas with insufficient amounts of sunlight and low vitamin D content in the diet. Besides, a recent systematic review and meta-analysis investigated a large amount of data indicating that the correlation between serum 25(OH)D levels and latitude was insignificant in regions of Russia, Ukraine, and Belarus (latitude  $45^\circ$ – $65^\circ$  N) [33]. On the basis of studies related to the correlation

between vitamin D status and latitude, it is possible to speculate that the geographical factor itself might lost its significance due to technological innovations, such as a fortified diet, the use of vitamin D preparations/supplements, and artificial sources of UV light. However, other studies indicated that depending on latitude, season or time of the day, actual exposure to natural UV light is an important factor determining vitamin cutaneous D production and vitamin D status [34]. Therefore, a number of factors that interfere with endogenous vitamin D synthesis, including lifestyle (indoor or outdoor), wearing inappropriate of clothes [35] [36], aging, in particular, age over 65 years, use of sunscreens, and skin pigmentation [34] should be also included in risk stratification for vitamin D deficiency.

The VDR is the receptor for  $25(OH)_2D$ , the key regulator of calcium absorption in the gut and bone mineralization. It is also is a transcription factor regulating the expression of genes, which mediate its biologic activity. The VDR in humans is encoded by the VDR gene [37]. The VDR is widely distributed in tissues that determines its broad range of actions of 1,25(OH)<sub>2</sub>D on many physiologic and pathologic processes, including bone mineral metabolism and BMD. The accumulating evidence suggests that the VDR polymorphisms may be accountable for different molecular and functional outcomes of the VDR gene. The genetic variation associated with VDR polymorphism and its influence on serum vitamin D levels and BMD were explored in different populations. The presence of the following alleles was considered the most interesting in terms of their impact on calcium absorption and BMD: located in exon 2, FokI (rs10735810) and located between exons 8 and 9, BsmI (rs1544410), ApaI (rs7975232) and TaqaI (rs731236) [38]. It was demonstrated that these VDR polymorphisms were not associated with BMD or with fractures [39]. Nevertheless, another study revealed that VDR gene FokI and BsmI polymorphism was considerably associated with low BMD in North Indian postmenopausal women with osteoporosis [40]. Using meta-analysis, the effect of VDR BsmI polymorphism on osteoporosis risk in postmenopausal women and Africans was also shown [41]. It was found that the b/b variant had a significant risk reduction of developing osteoporosis in these cohorts of subjects, compared to Caucasians and Asians, suggesting that this genotype may protect individuals against the development of osteoporosis. Likewise, another meta-analysis presented evidence indicating that VDR BsmI B/b gene variant was not associated with the susceptibility of osteoporosis in the overall population, Caucasians, and Asians [42]. Moreover, the accumulated data suggested that the functional outcomes of genetic variations of the VDR gene in the pathogenesis of osteoporosis can be affected by regional and ethnic factors. The association of FokI, BsmI, ApaI, and TaqaI VDR polymorphisms with the features of growth and development of bone tissue was shown in populations of different origin, including Caucasian, African, and Asian [43] [44] [45] [46]. It was found that African-Americans and ethnic whites with the same VDR genotype differed in bone mass [47]. Peak bone mass is pivotal in the osteoporosis developmental process because it is considered as a hallmark marker of bone health, which can be affected by not only by genetic factors but also by nutritional, mechanical, and hormonal factors [48]. The physiological mechanism of the effect of VDR genotype polymorphism on BMD remains not fully understood. In groups of Asian origin, the VDR \* G BsmI allele was associated with a protective effect on the status of bone tissue, while in Caucasians, its effect was close to the risk of osteoporosis development [49]. In subjects of Central and Northern Europe populations, Russian including, the association between VDR FokI, BsmI, and TaqaI polymorphisms and bone tissue content was different from that described in subjects from Southern European regions [43]. Remarkably, the association of the VDR genotype with bone tissue status was established in groups, which are close anthropologically (Asians, Africans, or Caucasians) and living under similar environmental conditions. In this way, the associations of the VDR genotypes with bone

turnover parameters found in populations of Central and Northern Russia were similar to those from the Netherlands, Sweden, and Poland, but differed from those from Turkey, Southern Italy, India, and Japan [43] [47]. We consider a methodologically important result the fact that the association between the *VDR* genotype with the status of bone tissue was reliably manifested in groups combined with the ecological parameters of the environment. It can be assumed that, in populations living in regions with different levels of UV insolation and different availability of vitamin D<sub>2</sub>-containing foods, variants of bone metabolism are evolutionarily acquired and are determined by the alleles localized in different parts of the *VDR* 

gene but with similar functional outcomes. Therefore, genetic testing for the polymorphism of the VDR gene should be included in osteoporosis risk stratification because VDR polymorphisms may serve as useful markers for osteoporosis screening in some ethnicities. Moreover, screening of these genetic markers may help early identification of risk groups so preventive measures can be applied in a timely fashion and also to improve the effectiveness of therapy, avoid complications, lower disability and mortality rates in these patients, as well as to reduce the treatment expenses.

In addition, there are other osteoporosis-related genes that can be identified for the susceptibility of osteoporosis. To date, more than 150 differentially expressed genes are known to be involved in bone metabolism and osteoporosis development [50]. Human genetic studies have found many candidate genes of high susceptibility, for example, *COL1A* (encoding collagen type I alpha chains), *ESR1* (encoding estrogen receptor 1), *IL6* (encoding interleukin 6), *LCT* (encoding lactase which hydrolyzes lactose to glucose and galactose in the small intestine), and *LRP5* (encoding a low-density lipoprotein transmembrane receptor), related to BMD, biochemical markers of bone turnover, and bone fractures [51] [52] [53] [54]. Despite the fact that the exact contribution of these genes to osteoporosis remains unclear and may vary between people of different ethnic groups, the detection of some high susceptibility genetic markers of osteoporosis development may help to identify those subjects, who are at risk for osteoporosis development. One should bear in mind that the identification of all osteoporosis development-related genes would be burdensome since a large amount of sequencing information should be obtained and accurately interpreted.

# 4. Nutritional Prevention of the Vitamin D deficiency and Osteoporosis

#### 4.1 Plant-Based Vitamin D compounds

Along with the evolutionary selection of advantageous *VDR* genotypes in those living in conditions of cholecalciferol  $D_3$  deficiency, the established food habits play an important role in the maintenance of healthy bone tissue metabolism. In populations living in high-latitude regions, the traditional diet is significant in the maintaining of the sufficient serum concentration of 25(OH)D, because vitamin D synthesis in the skin is unlikely to occur during a substantial part of the year in these regions [55] [56] [57] [58] [59] [60]. In populations of high latitude, components of the "northern diet", such as fish (marine and, to a lesser extent, from local freshwater) [55] [56], reindeer venison and fat [57] [58], as well as meat and fat of marine mammals [59] [60] are accountable for the protective effects against vitamin D hypovitaminosis. However, the specificity and locality of these foods (except for fish) limit recommendations for their wide consumption to prevent vitamin D deficits. Therefore, it is important to identify alternative foods of plant origin that would provide sufficient dietary or fortification source of vitamin D compounds for the populations of temperate and southern regions without provoking criticism from devotees of some religious beliefs and cultural

traditions. In this regard, yeast, algae, lichens, and mushrooms have attracted special attention. The consumption of these foods may help to accommodate dietary diversity in vitamin D-rich foods. Improving the dietary intake of vitamin D is a nutritional target in vitamin D deficiency prevention.

The UV-treated baker's yeast can accumulate an enhanced content of vitamin  $D_2$  [61]. The addition of UV-treated baker's yeast to bread products exhibited a positive effect on vitamin D status comparable with using pure vitamin  $D_2$  [61]. For that reason, the use of UV-treated yeast for baking has been approved by the Food and Drug Administration and by the European Food Safety Authority, in the United States and Europe, respectively [61] [62].

Being a part of phytoplankton, algae and cyanobacteria, the ultimate producers of ergo- and cholecalciferol, are the promising sources of vitamin D [63]. In nature, planktivorous fish accumulates both vitamins  $D_2$  and  $D_3$  from planktonic food. At the next stage of the food chain, the consumption of planktivorous fish by larger animals (predatory fish, marine mammals, or humans) can support their vitamin D status. However, in aquaculture, feeding fish with food without or in small quantities of natural algae leads to the low accumulation of vitamin D compounds in fish [64]. Therefore, the inclusion of ergo- and calciferol-containing algae and cyanobacteria in the fish production cycle can be beneficial for vitamin D status in humans. The occurrence of vitamin  $D_3$  in algae indicates that vitamin  $D_3$  may be existent in other plants. Vitamin  $D_3$  has been identified in several plant species, including species of the Solanaceae family [65]. This family includes broadly distributed and consumed vegetables, such as potatoes, tomatoes, and pepper, which also have been found to contain vitamin  $D_3$ . Even though vitamin  $D_3$  was found in non-edible leaves of these plants rather than the edible portions, such as tubers or fruit (**Table 1**), these leaves should be considered to be utilized for manufacture of natural food supplements.

Moreover, lichens can be taken into consideration as a potential source of both vitamin  $D_2$  and  $D_3$  since these compounds were identified in the thallus of a lichen species in significant quantities [66]. In the thalli of *Cladina arbuscula*, the content of vitamin  $D_3$  was found ranging between 67 µg and 204 µg per 100 g of dry weight, and vitamin  $D_2$  content was between 22 µg and 55 µg per 100 g of dry weight [66]. In these amounts, lichens are superior in vitamin D content to traditionally recommended animal products, such as sea fish of fatty varieties (salmon and herring) [67] [68].

Under optimal growing conditions (i.e., optimal temperature, UV dosage, and moisture content), some edible mushrooms can serve as a source of ergocalciferol [69] [70] [71] [72]. In shiitake mushrooms, the amount of vitamin D<sub>2</sub> was found to be high  $29.87 \pm 1.38 \mu g/g$  (dry weight) [73]. In UV-irradiated oyster mushrooms, the content of vitamin D<sub>2</sub> increased by a maximum of 204.7  $\mu g/g$  [74]. Importantly, the remarkable bioavailability of vitamin D compounds in mushrooms was demonstrated [70]. This study showed that the intake of 2000 IU of vitamin D<sub>2</sub> contained in mushrooms was just as effective as taking of 2000 IU of vitamin D concentrations. Nonetheless, the use of mushrooms as potential sources of vitamin D requires further investigation, including population genetic studies. According to preliminary data, 2%-6% of the population of the European part of the Russian Federation, as well as 7%-30% of populations of Siberia and the Far East carry the AA\*TREH genotype responsible for the poor absorption of fungi [75] [76].

Summarized data of plant-based sources of vitamin D are presented in **Table 1**. The availability of organic food sources of vitamin D is essential in vitamin D deficiency prevention. It is worth mentioning that plant-based sources of vitamin D may also contain naturally occurring polyphenols that possess properties modulating bone metabolism. This is

applicable to potatoes, capsicum, tomatoes, yeast, algae, lichens, mushrooms, goji berries, and soybeans [77]. The protective effects of polyphenols on bone metabolism will be discussed in the next subsection.

**Table 1.** Plant-Based Sources of Vitamin D: Content and Availability.

Plant name/Botanical name	Vitamin D content (µg/100g)	Availability
Australian seaweeds [78]	0.03-0.67	High
Common chanterelles ( <i>Cantharellus cibarius</i> ) [79] [80]	5.2-28.1	High
Shiitake mushrooms (Lentinula edodes) [81]	22-110	High availability in
		Southeast Asia
Morel edible (Morchella esculenta) [79]	5.2-28.1	High
Sun-treated oyster mushrooms ( <i>Pleurotus ostreatus</i> ) [82].	67.4	High
Common miller (Lactarius trivialis) [70]	29	High
Romaine Lettuce (Lactuca sativa L. var. longifolia) [83]	9.5	High
Tomato leaves (Lycopersicon esculentum) [84] [85]	110	High
Potato leaves (Solanum tuberosum) [84]	15	High
Zucchini leaves ( <i>Cucurbita pepo</i> ) [84]	23	High
Pepper ( <i>Capsicum annuum L.</i> ) [85]	0.29-0.63	High
Reygras (Lolium perenne) [66]	0.07-6.4	High
Baker's yeast [61]	0.5	High
Wheat germ oil [86]	22.1-34.2	High
Avocado oil [86]	4.2-23.4	High
Sunflower oil [86]	7.9-17.4	High
Rapeseed oil [86]	4.1-9.5	High
Linseed oil [86]	4.1-9.5	High
Olive oil <b>[86]</b>	4.5	High
Common pumpkin ( <i>Cucurbita moschata</i> ) [63]	23	High
Sowing alfalfa (Medicago sativa) [63]	0.062-0.1	High
Triseta yellowish [63]	10	High
Tobacco tree [63]	30-100	High
Lichen (Cladina arbuscula) [66]	67-204	High
Microalgae [63]	5.0-15	Average
White mushrooms [72]	58.7	Average
Double champignons (Agaricus bisporus) [79]	0.1-0.3	Average
Goji berries [87]	0.90	Average
Fresh kombu (Lessonia corrugate) [78]	0.01	Low
Danish champignons (Agaricus bisporus) [88]	17.6	Low
Maitake (Grifola frondosa) [79]	5.2-28.1	Low
Japanese wireweed (Sargassum muticum) [78]	90	Low

4.2 Other Plant-Derived Compounds Influencing Bone Metabolism

Some nutrigenomics data suggested that the expression and functional activity of osteoporosis-related genes can be positively influenced by edible plant-derived compounds [89]. This study showed that the expression of osteoblastic genes (alkaline phosphatase and osteocalcin), were increased by calophyllolide, the main component of the Alexandria laurel (*Calophylluminophyllum*) of the Clusia family (*Clusiaceae*). Calophyllolide also induced differentiation of osteoblasts in murine osteoblastic cells. Osteoblast differentiation is important for maintaining appropriate bone rigidity, strength, and, to some extent, elasticity. Pomegranate (*Punica granatum*) also increased osteoblast differentiation and the level of the *Runx2* gene expression in osteoblasts [90]. Unfortunately, the amount of evidence that could explain the protective effects of edible plants by their influence on the expression of disease-associated genes is scarce.

Most of the available data linking the protective effects of plant-based dietary components with osteoporosis development is either empirical or accompanied by biochemical explanations. It was found that foods containing estrogens (for example, soybean or flaxseed), as well as polyunsaturated fatty acids (linolenic, eicosapentaenoic, and docosahexaenoic acids) favorably affected the age-related development of bone tissue [91] [92] [93]. In particular, metabolized by intestinal microflora from secoisolariciresinol diglycoside and matairesinol, mammalian lignans (found in flaxseeds), enterodiol and enterolactone are believed to protect from osteoporosis [89]. They have improved in vitro cell viability, DNA content, alkaline phosphatase activity, and the expression of genetic markers, which modulate bone formation (osteonectin and type 1 collagen). Phytoestrogens, a diverse group of naturally occurring nonsteroidal plant compounds, are generally considered to prevent osteoporosis by promoting bone health. Phytoestrogens represent a group of compounds, which include isoflavones (genistein, daidzein, and equol), lignans (enterolactone and enterodiol), coumestans (coumestrol), stilbenes (resveratrol), and flavonoids (quercetin and kaempherol). They resemble estradiol in the molecular structure and by binding high affinity estrogen receptors display estradiol-like effects. In particular, soy isoflavones were demonstrated to act on both osteoblasts and osteoclasts via genomic and non-genomic pathways and, thus, ensure beneficial effects on BMD, bone turnover markers, and bone mechanical strength in postmenopausal women or in women in the early postnatal period [93] [94] [95]. Similarly, a mixture of flavonoid-containing extracts of Herba epimedii and Fructus ligustri lucidi that are abundantly occurred in China was shown to be able to rebalance bone remodeling in women with postmenopausal osteoporosis [96]. Additionally, resveratrol produced in the skin of grapes in high quantities exhibited osteoprotective and chondroprotective properties [97] and, as supported by preclinical evidence from rat models of osteoporosis, it may be applied as a therapeutic agent for bone loss [98]. Noteworthy, the main food sources of resveratrol are wine, berries, peanuts, and soy [99].

Relying on the evidence indicating a close link between factors involved in inflammation and those essential for bone physiology and remodeling [100] [101], it was proposed that the maladaptive link between inflammation and bone turnover may be a major determinant of osteoporosis [102]. Nutritional plant-based components can determine this link. The essential oils and monoterpene of thyme and rosemary were shown to be effective inhibitors of bone resorption and inflammation [82]. Furthermore, found in many fruit, vegetables, cereals, and beverages, plant polyphenols are the most studied compounds, which exhibited a positive impact on bone metabolism. Polyphenols can preserve bone health by the following actions: (i) by diminishing bone loss via anti-inflammatory action; (ii) by reducing bone loss via antioxidant activity; (iii) by improving osteoblastogenesis; (iv) by reducing osteoclastogenesis, and (v) by osteoimmunological activity [103]. In particular, the immune modulation effect of polyphenols is supported by their impact on populations of immune cells, proinflammatory cytokine synthesis, and gene expression, as reviewed in [104]. For example, polyphenolic compounds inhibited interleukin 6 (IL-6)-mediated inflammation by direct inhibition of the signal transduction pathway [105]. Notably, IL-6 mediated- inflammation is deeply implicated in ageing and age-related diseases, including osteoporosis [97]. An early study showed that IL-6-mediated inflammation can contribute to the process of bone remodeling by stimulating osteoclastogenesis and osteoclast activity [106]. The role of polyphenols in mitigating the damaging effects of reactive oxygen species-induced oxidative stress implicated in osteoporosis pathogenesis was also described [107]. The studies mentioned in this subsection elucidated the mechanisms determining the protective effects of naturally occurring phytoestrogens, flavonoids, and polyphenols on bone metabolism. Their effects are attributed to the ability to increase bone formation and reduce bone resorption. In this way, the intake of natural foods containing phytoestrogens, flavonoids, and polyphenols can be potentially recommended for the nutritional prevention of osteoporosis. However, the number of clinical studies indicating that the experimental data can be extrapolated to humans is limiting.

# 5. Concluding Remarks and Future Perspectives

The effective prevention of vitamin D deficiency and osteoporosis can be achieved with the timely identification of individuals with an increased risk and appropriate dietary intervention. Vitamin D deficiency and osteoporosis risk stratification should be complex and rely on not only the evidence indicating the impact of clinical risk factors and epidemiological data but also on the evidence specifying regional anthropological features of bone mass formation, as well as genetic, and nutrigenomic data. Overall, the following factors should be considered: (i) UV exposure determined by latitude, season, time of the day, lifestyle (indoor or outdoor), wearing protective clothes and sunscreens, and skin pigmentation; (ii) the local food habits; (iii) the accessibility of natural foods containing vitamin D, polyphenols, and other bone metabolism modulating compounds; (iv) ethnicity and *VDR* genotype since the *VDR* genotype that can determine a specific bone metabolism in different populations. At present, the associations between the *VDR* genotype and bone metabolism are poorly understood. It can be assumed that these associations are evolutionally acquired and differ in populations of different origin and living under different environmental conditions.

The current evidence suggests that dietary intervention with natural foods may be helpful to prevent vitamin D deficiency and osteoporosis development. However, further large-scale human clinical and epidemiological studies are required to establish a clear relationship between the consumption of plant-based compounds influencing bone health. It can be anticipated that the effective levels of intake will be determined in the near future for individuals at risk of developing osteoporosis. Moreover, long-term human intervention studies are needed, in which, in addition to protective effects on the bone, safety aspects will be evaluated. Finally, the identification of novel genes and molecular pathways will potentially help to develop novel prevention and therapeutic strategies for patients with osteoporosis.

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# References

- Munns, C.F.; Shaw, N.; Kiely, M.; Specker, B.L.; Thacher, T.D.; Ozono, K.; Michigami, T.; Tiosano, D.; Mughal, M.Z.; Mäkitie, O.; et al. Global Consensus Recommendations on Prevention and Management of Nutritional Rickets. *HRP* 2016, 85, 83–106, doi:10.1159/000443136.
- Rizzoli, R.; Boonen, S.; Brandi, M.-L.; Bruyère, O.; Cooper, C.; Kanis, J.A.; Kaufman, J.-M.; Ringe, J.D.; Weryha, G.; Reginster, J.-Y. Vitamin D Supplementation in Elderly or Postmenopausal Women: A 2013 Update of the 2008 Recommendations from the European Society for Clinical and Economic Aspects of Osteoporosis and Osteoarthritis (ESCEO). *Curr Med Res Opin* 2013, *29*, 305–313, doi:10.1185/03007995.2013.766162.
- Bischoff-Ferrari, H.A.; Kiel, D.P.; Dawson-Hughes, B.; Orav, J.E.; Li, R.; Spiegelman, D.; Dietrich, T.; Willett, W.C. Dietary Calcium and Serum 25-Hydroxyvitamin D Status in Relation to BMD among U.S. Adults. *J. Bone Miner. Res.* 2009, 24, 935–942, doi:10.1359/jbmr.081242.
- 4. Bischoff-Ferrari, H.A.; Willett, W.C.; Wong, J.B.; Stuck, A.E.; Staehelin, H.B.; Orav, E.J.; Thoma, A.; Kiel, D.P.; Henschkowski, J. Prevention of Nonvertebral Fractures With Oral Vitamin D and Dose Dependency: A Meta-Analysis of Randomized Controlled Trials. *Arch Intern Med* **2009**, *169*, 551, doi:10.1001/archinternmed.2008.600.
- Ja, C.; Az, L.; L, W.; M, H.; Me, D.; Dc, B.; Js, L.; Rd, J.; Ja, R.; C, W.; et al. Serum 25-Hydroxyvitamin D Concentrations and Risk for Hip Fractures. *Ann Intern Med* 2008, *149*, 242–250, doi:10.7326/0003-4819-149-4-200808190-00005.
- 6. Koo, W.; Walyat, N. Vitamin D and Skeletal Growth and Development. *Curr Osteoporos Rep* **2013**, *11*, 188–193, doi:10.1007/s11914-013-0156-1.
- Kadam, N.S.; Chiplonkar, S.A.; Khadilkar, A.V.; Khadilkar, V.V. Prevalence of Osteoporosis in Apparently Healthy Adults above 40 Years of Age in Pune City, India. *Indian J Endocrinol Metab* 2018, 22, 67–73, doi:10.4103/ijem.IJEM\_438\_17.
- 8. Campion, J.M.; Maricic, M.J. Osteoporosis in Men. AFP 2003, 67, 1521–1526.
- Ahmed, S.F.; Elmantaser, M. Secondary Osteoporosis. *Endocr Dev* 2009, 16, 170–190, doi:10.1159/000223695.
- Zamani, M.; Zamani, V.; Heidari, B.; Parsian, H.; Esmaeilnejad-Ganji, S.M. Prevalence of Osteoporosis with the World Health Organization Diagnostic Criteria in the Eastern Mediterranean Region: A Systematic Review and Meta-Analysis. *Arch Osteoporos* 2018, *13*, 129, doi:10.1007/s11657-018-0540-7.
- 11. Lindsay, R.; Burge, R.T.; Strauss, D.M. One Year Outcomes and Costs Following a Vertebral Fracture. *Osteoporos Int* **2005**, *16*, 78–85, doi:10.1007/s00198-004-1646-x.
- 12. van Schoor, N.M.; Visser, M.; Pluijm, S.M.F.; Kuchuk, N.; Smit, J.H.; Lips, P. Vitamin D Deficiency as a Risk Factor for Osteoporotic Fractures. *Bone* **2008**, *42*, 260–266, doi:10.1016/j.bone.2007.11.002.

- Snijder, M.B.; van Schoor, N.M.; Pluijm, S.M.F.; van Dam, R.M.; Visser, M.; Lips, P. Vitamin D Status in Relation to One-Year Risk of Recurrent Falling in Older Men and Women. *The Journal of Clinical Endocrinology & Metabolism* 2006, *91*, 2980–2985, doi:10.1210/jc.2006-0510.
- 14. Johnell, O.; Kanis, J.A. An Estimate of the Worldwide Prevalence and Disability Associated with Osteoporotic Fractures. *Osteoporos Int* **2006**, *17*, 1726–1733, doi:10.1007/s00198-006-0172-4.
- Wright, N.C.; Looker, A.C.; Saag, K.G.; Curtis, J.R.; Delzell, E.S.; Randall, S.; Dawson-Hughes, B. The Recent Prevalence of Osteoporosis and Low Bone Mass in the United States Based on Bone Mineral Density at the Femoral Neck or Lumbar Spine. *Journal of Bone and Mineral Research* 2014, 29, 2520–2526, doi:10.1002/jbmr.2269.
- Sanders, K.M.; Stuart, A.L.; Williamson, E.J.; Simpson, J.A.; Kotowicz, M.A.; Young, D.; Nicholson, G.C. Annual High-Dose Oral Vitamin D and Falls and Fractures in Older Women: A Randomized Controlled Trial. *JAMA* 2010, *303*, 1815–1822, doi:10.1001/jama.2010.594.
- Salovaara, K.; Tuppurainen, M.; Kärkkäinen, M.; Rikkonen, T.; Sandini, L.; Sirola, J.; Honkanen, R.; Alhava, E.; Kröger, H. Effect of Vitamin D3 and Calcium on Fracture Risk in 65- to 71-Year-Old Women: A Population-Based 3-Year Randomized, Controlled Trial—the OSTPRE-FPS. *Journal of Bone and Mineral Research* 2010, 25, 1487–1495, doi:https://doi.org/10.1002/jbmr.48.
- 18. Lips, P.; van Schoor, N.M. The Effect of Vitamin D on Bone and Osteoporosis. *Best Practice & Research Clinical Endocrinology & Metabolism* **2011**, *25*, 585–591, doi:10.1016/j.beem.2011.05.002.
- 19. (PDF) Vitamin D and Health: Evolution, Biologic Functions, and Recommended Dietary Intakes for Vitamin D Available online: https://www.researchgate.net/publication/226676251\_Vitamin\_D\_and\_Health\_Evolutio n\_Biologic\_Functions\_and\_Recommended\_Dietary\_Intakes\_for\_Vitamin\_D (accessed on 30 September 2020).
- 20. Ložnjak, P.; Jakobsen, J. Stability of Vitamin D3 and Vitamin D2 in Oil, Fish and Mushrooms after Household Cooking. *Food Chemistry* **2018**, *254*, 144–149, doi:10.1016/j.foodchem.2018.01.182.
- 21. Holick, M.F. Vitamin D: Evolutionary, Physiological and Health Perspectives. *Curr Drug Targets* **2011**, *12*, 4–18, doi:10.2174/138945011793591635.
- 22. Lips, P. Vitamin D Deficiency and Secondary Hyperparathyroidism in the Elderly: Consequences for Bone Loss and Fractures and Therapeutic Implications. *Endocrine Reviews* **2001**, *22*, 477–501, doi:10.1210/edrv.22.4.0437.
- 23. Holick, M.F. Vitamin D Deficiency. *N Engl J Med* **2007**, *357*, 266–281, doi:10.1056/NEJMra070553.
- 24. Pouresmaeili, F.; Kamalidehghan, B.; Kamarehei, M.; Goh, Y.M. A Comprehensive Overview on Osteoporosis and Its Risk Factors. *Ther Clin Risk Manag* **2018**, *14*, 2029–2049, doi:10.2147/TCRM.S138000.
- Pfeifer, M.; Begerow, B.; Minne, H.W.; Schlotthauer, T.; Pospeschill, M.; Scholz, M.; Lazarescu, A.D.; Pollähne, W. Vitamin D Status, Trunk Muscle Strength, Body Sway, Falls, and Fractures among 237 Postmenopausal Women with Osteoporosis. *Exp Clin Endocrinol Diabetes* 2001, 109, 87–92, doi:10.1055/s-2001-14831.
- 26. Tsai, K.S.; Hsu, S.H.; Cheng, J.P.; Yang, R.S. Vitamin D Stores of Urban Women in Taipei: Effect on Bone Density and Bone Turnover, and Seasonal Variation. *Bone* **1997**, 20, 371–374, doi:10.1016/s8756-3282(97)00010-0.

- Hillier, T.A.; Stone, K.L.; Bauer, D.C.; Rizzo, J.H.; Pedula, K.L.; Cauley, J.A.; Ensrud, K.E.; Hochberg, M.C.; Cummings, S.R. Evaluating the Value of Repeat Bone Mineral Density Measurement and Prediction of Fractures in Older Women: The Study of Osteoporotic Fractures. *Archives of Internal Medicine* 2007, 167, 155–160, doi:10.1001/archinte.167.2.155.
- 28. Bhan, A.; Rao, A.D.; Rao, D.S. Osteomalacia as a Result of Vitamin D Deficiency. *Endocrinology and Metabolism Clinics* **2010**, *39*, 321–331, doi:10.1016/j.ecl.2010.02.001.
- 29. Tiosano, D.; Hochberg, Z. Hypophosphatemia: The Common Denominator of All Rickets. *J Bone Miner Metab* **2009**, 27, 392–401, doi:10.1007/s00774-009-0079-1.
- 30. Davies, J.H.; Shaw, N.J. Preventable but No Strategy: Vitamin D Deficiency in the UK. *Arch. Dis. Child.* **2011**, *96*, 614–615, doi:10.1136/adc.2010.191627.
- 31. Nesse, R.M.; Bergstrom, C.T.; Ellison, P.T.; Flier, J.S.; Gluckman, P.; Govindaraju, D.R.; Niethammer, D.; Omenn, G.S.; Perlman, R.L.; Schwartz, M.D.; et al. Making Evolutionary Biology a Basic Science for Medicine. 8.
- 32. Kozlov, A.I., Vershubskaya, G. D-vitamin status and lactase persistence in European populations (literature review with meta-analysis elements). Moscow University Bulletin. Series 23: *Anthropology*. **2017**, *3*, 68-75. Read in Russian.
- 33. Kozlov, A.I.; Vershubsky, G.G. Blood Serum 25-Hydroxyvitamin D in Various Populations of Russia, Ukraine, and Belarus: A Systematic Review with Elements of Meta-Analysis. *Hum Physiol* **2017**, *43*, 729–740, doi:10.1134/S0362119717060044.
- 34. (PDF) Vitamin D and Health: Evolution, Biologic Functions, and Recommended Dietary Intakes for Vitamin D Available online: https://www.researchgate.net/publication/226676251\_Vitamin\_D\_and\_Health\_Evolutio n\_Biologic\_Functions\_and\_Recommended\_Dietary\_Intakes\_for\_Vitamin\_D (accessed on 30 September 2020).
- Alagöl, F.; Shihadeh, Y.; Boztepe, H.; Tanakol, R.; Yarman, S.; Azizlerli, H.; Sandalci, Ö. Sunlight Exposure and Vitamin D Deficiency in Turkish Women. *J Endocrinol Invest* 2000, 23, 173–177, doi:10.1007/BF03343702.
- 36. Ardawi, M.-S.M.; Sibiany, A.M.; Bakhsh, T.M.; Qari, M.H.; Maimani, A.A. High Prevalence of Vitamin D Deficiency among Healthy Saudi Arabian Men: Relationship to Bone Mineral Density, Parathyroid Hormone, Bone Turnover Markers, and Lifestyle Factors. *Osteoporos Int* 2012, 23, 675–686, doi:10.1007/s00198-011-1606-1.
- Szpirer, J.; Szpirer, C.; Riviere, M.; Levan, G.; Marynen, P.; Cassiman, J.-J.; Wiese, R.; DeLuca, H.F. The Sp1 Transcription Factor Gene (SP1) and the 1,25-Dihydroxyvitamin D3 Receptor Gene (VDR) Are Colocalized on Human Chromosome Arm 12q and Rat Chromosome 7. *Genomics* 1991, *11*, 168–173, doi:10.1016/0888-7543(91)90114-T.
- Uitterlinden, A.G.; Fang, Y.; van Meurs, J.B.J.; Pols, H.A.P.; van Leeuwen, J.P.T.M. Genetics and Biology of Vitamin D Receptor Polymorphisms. *Gene* 2004, *338*, 143–156, doi:10.1016/j.gene.2004.05.014.
- Uitterlinden, A.G.; Ralston, S.H.; Brandi, M.L.; Carey, A.H.; Grinberg, D.; Langdahl, B.L.; Lips, P.; Lorenc, R.; Obermayer-Pietsch, B.; Reeve, J.; et al. The Association between Common Vitamin D Receptor Gene Variations and Osteoporosis: A Participant-Level Meta-Analysis. *Annals of Internal Medicine* 2006, 145, 255–264, doi:10.7326/0003-4819-145-4-200608150-00005.
- 40. Ahmad, I.; Jafar, T.; Mahdi, F.; Arshad, Md.; Das, S.K.; Waliullah, S.; Mahdi, A.A. Association of Vitamin D Receptor (FokI and BsmI) Gene Polymorphism with Bone Mineral Density and Their Effect on 25-Hydroxyvitamin D Level in North Indian

Postmenopausal Women with Osteoporosis. *Indian J Clin Biochem* **2018**, *33*, 429–437, doi:10.1007/s12291-017-0706-x.

- Jia, F.; Sun, R.-F.; Li, Q.-H.; Wang, D.-X.; Zhao, F.; Li, J.-M.; Pu, Q.; Zhang, Z.-Z.; Jin, Y.; Liu, B.-L.; et al. Vitamin D Receptor BsmI Polymorphism and Osteoporosis Risk: A Meta-Analysis from 26 Studies. *Genetic Testing and Molecular Biomarkers* 2012, *17*, 30–34, doi:10.1089/gtmb.2012.0267.
- 42. Qin, G.; Dong, Z.; Zeng, P.; Liu, M.; Liao, X. Association of Vitamin D Receptor BsmI Gene Polymorphism with Risk of Osteoporosis: A Meta-Analysis of 41 Studies. *Mol Biol Rep* **2013**, *40*, 497–506, doi:10.1007/s11033-012-2086-x.
- 43. Kozlov, A.I.; Vershubskaya, G.G.; Negasheva, M.A. Association between Relative Bone Mass and Vitamin D Receptor Gene Polymorphism. *Hum Physiol* **2017**, *43*, 320–325, doi:10.1134/S0362119717030100.
- 44. Zmuda, J.M.; Cauley, J.A.; Danielson, M.E.; Wolf, R.L.; Ferrell, R.E. Vitamin D Receptor Gene Polymorphisms, Bone Turnover, and Rates of Bone Loss in Older African-American Women. *Journal of Bone and Mineral Research* **1997**, *12*, 1446–1452, doi:10.1359/jbmr.1997.12.9.1446.
- 45. Rao Vupputuri, M.; Goswami, R.; Gupta, N.; Ray, D.; Tandon, N.; Kumar, N. Prevalence and Functional Significance of 25-Hydroxyvitamin D Deficiency and Vitamin D Receptor Gene Polymorphisms in Asian Indians. *Am J Clin Nutr* **2006**, *83*, 1411–1419, doi:10.1093/ajcn/83.6.1411.
- Kozlov, A.I.; Vershubskaya, G.G.; Ateeva, Yu.A.; Orr, P.; Larcombe, L. Association of Vitamin D Receptor Gene with Anthropometric Measures in Komi Ethnic Group. *Russ J Genet Appl Res* 2014, *4*, 397–404, doi:10.1134/S2079059714050074.
- 47. Nelson, D.A.; Vande Vord, P.J.; Wooley, P.H. Polymorphism in the Vitamin D Receptor Gene and Bone Mass in African-American and White Mothers and Children: A Preliminary Report. *Ann. Rheum. Dis.* **2000**, *59*, 626–630, doi:10.1136/ard.59.8.626.
- 48. Hamerman, D. Bone Health across the Generations: A Primer for Health Providers Concerned with Osteoporosis Prevention. *Maturitas* **2005**, *50*, 1–7, doi:10.1016/j.maturitas.2004.08.009.
- 49. Zintzaras, E.; Rodopoulou, P.; Koukoulis, G.N. BsmI, TaqI, ApaI and FokI Polymorphisms in the Vitamin D Receptor (VDR) Gene and the Risk of Osteoporosis: A Meta-Analysis. *Disease Markers* **2006**, *22*, 317–326.
- 50. Hopwood, B.; Tsykin, A.; Findlay, D.M.; Fazzalari, N.L. Gene Expression Profile of the Bone Microenvironment in Human Fragility Fracture Bone. *Bone* **2009**, *44*, 87–101, doi:10.1016/j.bone.2008.08.120.
- Bardai, G.; Moffatt, P.; Glorieux, F.H.; Rauch, F. DNA Sequence Analysis in 598 Individuals with a Clinical Diagnosis of Osteogenesis Imperfecta: Diagnostic Yield and Mutation Spectrum. *Osteoporos Int* 2016, 27, 3607–3613, doi:10.1007/s00198-016-3709-1.
- 52. Chantarangsu, S.; Sura, T.; Mongkornkarn, S.; Donsakul, K.; Torrungruang, K. Vitamin D Receptor Gene Polymorphism and Smoking in the Risk of Chronic Periodontitis. *Journal of Periodontology* **2016**, *87*, 1343–1351, doi:10.1902/jop.2016.160222.
- Alharbi, O.; El-Sohemy, A. Lactose Intolerance (LCT-13910C>T) Genotype Is Associated with Plasma 25-Hydroxyvitamin D Concentrations in Caucasians: A Mendelian Randomization Study. J Nutr 2017, 147, 1063–1069, doi:10.3945/jn.116.246108.
- 54. Zimmermann, A.; Popp, R.A.; Rossmann, H.; Bucerzan, S.; Nascu, I.; Leucuta, D.; Weber, M.M.; Grigorescu-Sido, P. Gene Variants of Osteoprotegerin, Estrogen-,

Calcitonin- and Vitamin D-Receptor Genes and Serum Markers of Bone Metabolism in Patients with Gaucher Disease Type 1. *Ther Clin Risk Manag* **2018**, *14*, 2069–2080, doi:10.2147/TCRM.S177480.

- 55. Brustad, M.; Sandanger, T.; Wilsgaard, T.; Aksnes, L.; Lund, E. Change in Plasma Levels of Vitamin D after Consumption of Cod-Liver and Fresh Cod-Liver Oil as Part of the Traditional North Norwegian Fish Dish "Molje." *International Journal of Circumpolar Health* **2003**, *62*, 40–53, doi:10.3402/ijch.v62i1.17527.
- 56. Kozlov, A.I.; Vershubsky, G.G. Systematic Review on 25-HydroxyvitaminD Levels in Various Populations of the Russian North. *Hum Physiol* **2019**, *45*, 565–575, doi:10.1134/S0362119719050062.
- 57. Wiklund, E.; Johansson, L. Water-Holding Capacity, Colour Stability and Sensory Characteristics in Meat (M. Longissimus Dorsi) from Reindeer Fed Two Different Commercial Feeds. *Rangifer* **2011**, *31*, 49–59, doi:10.7557/2.31.1.2019.
- Kozlov, A.; Khabarova, Y.; Vershubsky, G.; Ateeva, Y.; Ryzhaenkov, V. Vitamin D Status of Northern Indigenous People of Russia Leading Traditional and "Modernized" Way of Life. *International Journal of Circumpolar Health* 2014, 73, 26038, doi:10.3402/ijch.v73.26038.
- 59. Brustad, M.; Alsaker, E.; Engelsen, O.; Aksnes, L.; Lund, E. Vitamin D Status of Middle-Aged Women at 65–71°N in Relation to Dietary Intake and Exposure to Ultraviolet Radiation. *Public Health Nutrition* **2004**, *7*, 327–335, doi:10.1079/PHN2003536.
- 60. Kuhnlein, H.V.; Barthet, V.; Farren, A.; Falahi, E.; Leggee, D.; Receveur, O.; Berti, P. Vitamins A, D, and E in Canadian Arctic Traditional Food and Adult Diets. *Journal of Food Composition and Analysis* **2006**, *19*, 495–506, doi:10.1016/j.jfca.2005.02.007.
- 61. Scientific Opinion on the Safety of Vitamin D-enriched UV-treated Baker's Yeast. *EFS2* **2014**, *12*, doi:10.2903/j.efsa.2014.3520.
- 62. Food and Drug Administration, HHS Food Additives Permitted for Direct Addition to Food for Human Consumption; Folic Acid. Final Rule. *Fed Regist* **2016**, *81*, 22176–22183.
- 63. Jäpelt, R.B.; Jakobsen, J. Vitamin D in Plants: A Review of Occurrence, Analysis, and Biosynthesis. *Front. Plant Sci.* **2013**, *4*, doi:10.3389/fpls.2013.00136.
- Bell, J.G.; Waagbø, R. Safe and Nutritious Aquaculture Produce: Benefits and Risks of Alternative Sustainable Aquafeeds. In *Aquaculture in the Ecosystem*; Holmer, M., Black, K., Duarte, C.M., Marbà, N., Karakassis, I., Eds.; Springer Netherlands: Dordrecht, 2008; pp. 185–225 ISBN 978-1-4020-6810-2.
- 65. Boland, R.; Skliar, M.; Curino, A.; Milanesi, L. Vitamin D Compounds in Plants. *Plant Science* **2003**, *164*, 357–369, doi:10.1016/S0168-9452(02)00420-X.
- 66. Wang, T.; Bengtsson, G.; Kärnefelt, I.; Björn, L.O. Provitamins and Vitamins D2 and D3 in Cladina Spp. over a Latitudinal Gradient: Possible Correlation with UV Levels. *Journal of Photochemistry and Photobiology B: Biology* **2001**, *62*, 118–122, doi:10.1016/S1011-1344(01)00160-9.
- 67. Jakobsen, J.; Smith, C.; Bysted, A.; Cashman, K.D. Vitamin D in Wild and Farmed Atlantic Salmon (Salmo Salar)—What Do We Know? *Nutrients* **2019**, *11*, 982, doi:10.3390/nu11050982.
- 68. Aro, T.L.; Larmo, P.S.; Bäckman, C.H.; Kallio, H.P.; Tahvonen, R.L. Fatty Acids and Fat-Soluble Vitamins in Salted Herring (Clupea Harengus) Products. J. Agric. Food Chem. 2005, 53, 1482–1488, doi:10.1021/jf0401221.
- 69. Simon, R.R.; Phillips, K.M.; Horst, R.L.; Munro, I.C. Vitamin D Mushrooms: Comparison of the Composition of Button Mushrooms (Agaricus Bisporus) Treated

Postharvest with UVB Light or Sunlight. J. Agric. Food Chem. 2011, 59, 8724–8732, doi:10.1021/jf201255b.

- Keegan, R.-J.H.; Lu, Z.; Bogusz, J.M.; Williams, J.E.; Holick, M.F. Photobiology of Vitamin D in Mushrooms and Its Bioavailability in Humans. *Dermato-Endocrinology* 2013, 5, 165–176, doi:10.4161/derm.23321.
- 71. Kamweru, P.K.; Tindibale, E.L. Vitamin D and Vitamin D from Ultraviolet-Irradiated Mushrooms (Review). *IJM* **2016**, *18*, doi:10.1615/IntJMedMushrooms.v18.i3.30.
- 72. Cardwell, G.; Bornman, J.F.; James, A.P.; Black, L.J. A Review of Mushrooms as a Potential Source of Dietary Vitamin D. *Nutrients* **2018**, *10*, 1498, doi:10.3390/nu10101498.
- 73. Won, D.J.; Kim, S.Y.; Jang, C.H.; Lee, J.S.; Ko, J.A.; Park, H.J. Optimization of UV Irradiation Conditions for the Vitamin D2-Fortified Shiitake Mushroom (Lentinula Edodes) Using Response Surface Methodology. *Food Sci Biotechnol* **2018**, *27*, 417–424, doi:10.1007/s10068-017-0266-0.
- 74. Huang, S.-J.; Lin, C.-P.; Tsai, S.-Y. Vitamin D2 Content and Antioxidant Properties of Fruit Body and Mycelia of Edible Mushrooms by UV-B Irradiation. *Journal of Food Composition and Analysis* **2015**, *42*, 38–45, doi:10.1016/j.jfca.2015.02.005.
- 75. Kozlov, A.I. [Carbohydrate-related nutritional and genetic risks of obesity for indigenous northerners]. *Vopr Pitan* **2019**, *88*, 5–16, doi:10.24411/0042-8833-2019-10001.
- 76. (PDF) Anthropological and Ecological Specificity of Polymorphism in Genes Related to Bone Tissue Metabolism (as Exemplified by the Shors People) Available online: https://www.researchgate.net/publication/335643187\_Anthropological\_and\_ecological\_ specificity\_of\_polymorphism\_in\_genes\_related\_to\_bone\_tissue\_metabolism\_as\_exemp lified\_by\_the\_Shors\_people (accessed on 30 September 2020).
- 77. Bhagwat, S.; Haytowitz, D.B.; Wasswa-Kintu, S. Documentation and User Guide. 12.
- Hughes, L.J.; Black, L.J.; Sherriff, J.L.; Dunlop, E.; Strobel, N.; Lucas, R.M.; Bornman, J.F. Vitamin D Content of Australian Native Food Plants and Australian-Grown Edible Seaweed. *Nutrients* 2018, 10, doi:10.3390/nu10070876.
- Phillips, K.M.; Ruggio, D.M.; Horst, R.L.; Minor, B.; Simon, R.R.; Feeney, M.J.; Byrdwell, W.C.; Haytowitz, D.B. Vitamin D and Sterol Composition of 10 Types of Mushrooms from Retail Suppliers in the United States. *J. Agric. Food Chem.* 2011, 59, 7841–7853, doi:10.1021/jf104246z.
- Teichmann, A.; Dutta, P.C.; Staffas, A.; Jägerstad, M. Sterol and Vitamin D2 Concentrations in Cultivated and Wild Grown Mushrooms: Effects of UV Irradiation. *LWT - Food Science and Technology* 2007, 40, 815–822, doi:10.1016/j.lwt.2006.04.003.
- 81. Mattila, P.; Lampi, A.-M.; Ronkainen, R.; Toivo, J.; Piironen, V. Sterol and Vitamin D2 Contents in Some Wild and Cultivated Mushrooms. *Food Chemistry* **2002**, *76*, 293–298, doi:10.1016/S0308-8146(01)00275-8.
- 82. (PDF) Impact of the Natural Resource of UVB on the Content of Vitamin D 2 in Oyster Mushroom (Pleurotus Ostreatus) under Subtropical Settings Available online: https://www.researchgate.net/publication/326760240\_Impact\_of\_the\_Natural\_Resource \_of\_UVB\_on\_the\_Content\_of\_Vitamin\_D\_2\_in\_Oyster\_Mushroom\_Pleurotus\_ostreatu s\_under\_Subtropical\_Settings (accessed on 30 September 2020).
- Viñas, P.; Bravo-Bravo, M.; López-García, I.; Hernández-Córdoba, M. Dispersive Liquid–Liquid Microextraction for the Determination of Vitamins D and K in Foods by Liquid Chromatography with Diode-Array and Atmospheric Pressure Chemical Ionization-Mass Spectrometry Detection. *Talanta* 2013, 115, 806–813, doi:10.1016/j.talanta.2013.06.050.

- 84. Aburjai, T.; Al-Khalil, S.; Abuirjeie, M. Vitamin D3 and Its Metabolites in Tomato, Potato, Egg Plant and Zucchini Leaves. *Phytochemistry* **1998**, *49*, 2497–2499, doi:10.1016/S0031-9422(98)00246-5.
- Jäpelt, R.B.; Silvestro, D.; Smedsgaard, J.; Jensen, P.E.; Jakobsen, J. Quantification of Vitamin D3 and Its Hydroxylated Metabolites in Waxy Leaf Nightshade (Solanum Glaucophyllum Desf.), Tomato (Solanum Lycopersicum L.) and Bell Pepper (Capsicum Annuum L.). *Food Chemistry* 2013, 138, 1206–1211, doi:10.1016/j.foodchem.2012.11.064.
- 86. Baur, A.C.; Brandsch, C.; König, B.; Hirche, F.; Stangl, G.I. Plant Oils as Potential Sources of Vitamin D. *Front. Nutr.* **2016**, *3*, doi:10.3389/fnut.2016.00029.
- 87. (PDF) The Study of Biological Effects of Different Geographical Origin Goji Berries in Rats with Alimentary Hypercholesterolemia Available online: https://www.researchgate.net/publication/339439044\_The\_study\_of\_biological\_effects\_ of\_different\_geographical\_origin\_goji\_berries\_in\_rats\_with\_alimentary\_hypercholester olemia (accessed on 30 September 2020).
- 88. Urbain, P.; Jakobsen, J. Dose–Response Effect of Sunlight on Vitamin D2 Production in Agaricus Bisporus Mushrooms. J. Agric. Food Chem. 2015, 63, 8156–8161, doi:10.1021/acs.jafc.5b02945.
- 89. Feng, J.; Shi, Z.; Ye, Z. Effects of Metabolites of the Lignans Enterolactone and Enterodiol on Osteoblastic Differentiation of MG-63 Cells. *Biological and Pharmaceutical Bulletin* **2008**, *31*, 1067–1070, doi:10.1248/bpb.31.1067.
- 90. Siddiqui, S.; Arshad, M. Osteogenic Potential of Punica Granatum through Matrix Mineralization, Cell Cycle Progression and Runx2 Gene Expression in Primary Rat Osteoblasts. *DARU J Pharm Sci* **2014**, *22*, 72, doi:10.1186/s40199-014-0072-7.
- 91. Brooks, J.D.; Ward, W.E.; Lewis, J.E.; Hilditch, J.; Nickell, L.; Wong, E.; Thompson, L.U. Supplementation with Flaxseed Alters Estrogen Metabolism in Postmenopausal Women to a Greater Extent than Does Supplementation with an Equal Amount of Soy. *Am J Clin Nutr* 2004, *79*, 318–325, doi:10.1093/ajcn/79.2.318.
- Coetzee, M.; Haag, M.; Kruger, M.C. Effects of Arachidonic Acid, Docosahexaenoic Acid, Prostaglandin E2 and Parathyroid Hormone on Osteoprotegerin and RANKL Secretion by MC3T3-E1 Osteoblast-like Cells. *The Journal of Nutritional Biochemistry* 2007, 18, 54–63, doi:10.1016/j.jnutbio.2006.03.002.
- Atmaca, A.; Kleerekoper, M.; Bayraktar, M.; Kucuk, O. Soy Isoflavones in the Management of Postmenopausal Osteoporosis. *Menopause* 2008, 15, 748–757, doi:10.1097/gme.0b013e31815c1e7f.
- 94. Reinwald, S.; Weaver, C.M. Soy Isoflavones and Bone Health: A Double-Edged Sword? *J. Nat. Prod.* **2006**, *69*, 450–459, doi:10.1021/np058104g.
- 95. Franke, A.A.; Halm, B.M.; Kakazu, K.; Li, X.; Custer, L.J. Phytoestrogenic Isoflavonoids in Epidemiologic and Clinical Research. *Drug Testing and Analysis* **2009**, *1*, 14–21, doi:10.1002/dta.12.
- 96. Chen, Y.; Li, X.; Tang, X.; Gao, Y.; Yu, P.; Xu, L.; Liu, R. Combined Extracts of Herba Epimedii and Fructus Ligustri Lucidi Rebalance Bone Remodeling in Ovariectomized Rats Available online: https://www.hindawi.com/journals/ecam/2019/1596951/ (accessed on 30 September 2020).
- 97. Cooley, J.; Broderick, T.L.; Al-Nakkash, L.; Plochocki, J.H. Effects of Resveratrol Treatment on Bone and Cartilage in Obese Diabetic Mice. *J Diabetes Metab Disord* **2015**, *14*, 10, doi:10.1186/s40200-015-0141-6.

- 98. Tou, J.C. Evaluating Resveratrol as a Therapeutic Bone Agent: Preclinical Evidence from Rat Models of Osteoporosis. *Annals of the New York Academy of Sciences* **2015**, *1348*, 75–85, doi:10.1111/nyas.12840.
- 99. Burns, J.; Yokota, T.; Ashihara, H.; Lean, M.E.J.; Crozier, A. Plant Foods and Herbal Sources of Resveratrol. *J. Agric. Food Chem.* **2002**, *50*, 3337–3340, doi:10.1021/jf0112973.
- 100. Lorenzo, J. Interactions between Immune and Bone Cells: New Insights with Many Remaining Questions. *J Clin Invest* **2000**, *106*, 749–752, doi:10.1172/JCI11089.
- 101. Arron, J.R.; Choi, Y. Bone versus Immune System. *Nature* **2000**, *408*, 535–536, doi:10.1038/35046196.
- 102. Yun, A.J.; Lee, P.Y. Maldaptation of the Link between Inflammation and Bone Turnover May Be a Key Determinant of Osteoporosis. *Medical Hypotheses* 2004, 63, 532–537, doi:10.1016/S0306-9877(03)00326-8.
- 103. Nicolin, V.; De Tommasi, N.; Nori, S.L.; Costantinides, F.; Berton, F.; Di Lenarda, R. Modulatory Effects of Plant Polyphenols on Bone Remodeling: A Prospective View From the Bench to Bedside. *Front. Endocrinol.* **2019**, *10*, doi:10.3389/fendo.2019.00494.
- 104. Yahfoufi, N.; Alsadi, N.; Jambi, M.; Matar, C. The Immunomodulatory and Anti-Inflammatory Role of Polyphenols. *Nutrients* **2018**, *10*, 1618, doi:10.3390/nu10111618.
- 105. The Interleukin-6 Inflammation Pathway from Cholesterol to Aging--Role of Statins, Bisphosphonates and Plant Polyphenols in Aging and Age-Related Diseases. - Abstract -Europe PMC Available online: https://europepmc.org/article/PMC/1845171 (accessed on 30 September 2020).
- 106. Manolagas, S.C.; Jilka, R.L. Bone Marrow, Cytokines, and Bone Remodeling. Emerging Insights into the Pathophysiology of Osteoporosis. *N. Engl. J. Med.* **1995**, *332*, 305–311, doi:10.1056/NEJM199502023320506.
- 107. Rao, L.G.; Rao, A.V. Oxidative Stress and Antioxidants in the Risk of Osteoporosis Role of Phytochemical Antioxidants Lycopene and Polyphenol-Containing Nutritional Supplements. *Phytochemicals - Isolation, Characterisation and Role in Human Health* 2015, doi:10.5772/60446.